

# Activated neutrophils degrade cartilage endplates

Irina Heggli<sup>\*1</sup>, Mohamed Habib<sup>2</sup>, Justin Scheer<sup>3</sup>, Nick Herger<sup>1</sup>, Tamara Mengis<sup>1</sup>, Borbala Aradi-Vegh<sup>1</sup>, Christoph J. Laux<sup>4</sup>, José Miguel Spirig<sup>4</sup>, Florian Wanivenhaus<sup>4</sup>, Michael Betz<sup>4</sup>, Christopher P. Ames<sup>3</sup>, Mazda Farshad<sup>4</sup>, Oliver Distler<sup>1</sup>, Aaron J. Fields<sup>2</sup>, Stefan Dudli<sup>1</sup>

<sup>1</sup>Center of Experimental Rheumatology, Balgrist Campus, University Hospital Zurich and Balgrist University Hospital, University of Zurich, Switzerland; <sup>2</sup>Department of Orthopaedic Surgery, University of California San Francisco, San Francisco, CA, USA, <sup>3</sup>Department of Neurological Surgery, University of California San Francisco, San Francisco, CA, USA, <sup>4</sup>Department of Orthopedics, Balgrist University Hospital, University of Zurich, Switzerland; irina.heggli@usz.ch

## Introduction

### Modic changes (MC):

- Painful vertebral bone marrow lesions that occur around a degenerated intervertebral disc (IVD) and colocalize with endplate damage.
- Signs for neutrophil contribution in MC<sup>1,2</sup>: Granulation tissue at bone-disc junction and dysregulated neutrophil maturation in MC bone marrow

In rheumatoid arthritis: Activated neutrophils mediate articular cartilage<sup>3</sup>

Little is known about the role of neutrophils in Modic changes and disc tissue damage

## Aims

- To show that MC1 bone marrow neutrophils are activated
- To discover the effects of activated blood neutrophils on cartilage endplate (CEP) composition

## Methods

### MC1 bone marrow derived neutrophil activation assessment

- Isolation of MC1 and intra-patient control bone marrow neutrophils from patients undergoing spinal fusion (n = 5+5)
- RNA isolation, sequencing, bioinformatic overrepresentation analysis (ORA) and gene set enrichment analysis (GSEA)
- Measurement of the neutrophil activation marker CD66b with flow cytometry (n = 3+3) (Figure 1)

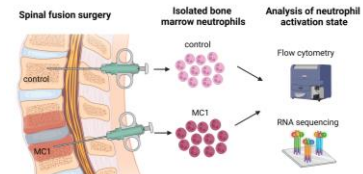


Figure 1. Bone marrow neutrophil collection and analysis.

### Neutrophil-mediated cartilage endplate (CEP) damage model

- Collection of 3 lumbar circular CEP biopsies per patient (patients n=6); CEP biopsies were halved and exposed to activated (100nM PMA, 3h) and non-activated neutrophil supernatant of one donor
- CEP tissue released sulphated glycosaminoglycan (sGAG) and hydroxyproline (as collagen release measure) were assayed using dimethylmethylene blue and chloramine-T assay (release from half-biopsy specific control was set to 100 %) (Figure 2)

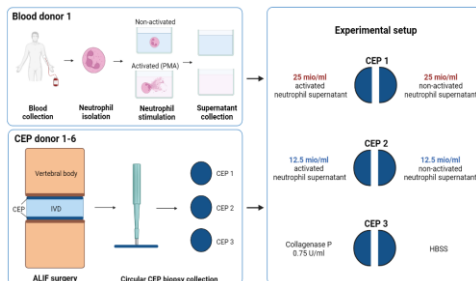


Figure 2. Neutrophil-mediated cartilage endplate (CEP) damage model.

## Results

### 1. MC1 bone marrow neutrophils have an activated pro-inflammatory phenotype

#### RNA sequencing:

- 185 differentially expressed genes between MC1 and control bone marrow neutrophils
- ORA: Top 5 upregulated gene ontology (GOs) in MC1: All related to neutrophil activation (Figure 3a).
- GSEA: Top enriched hallmark gene sets in MC1: "inflammatory response" (p=0.001, normalized enrichment score (NES) =1.6) (Figure 3b), "IFN-α response (p=0.0, NES=2.2), and IFN-γ response (p=0.0, NES=1.9)

#### Flow cytometric analysis of CD66b expression:

- Significantly more activated neutrophils (measured as CD66<sup>high</sup> expression) in MC1 (control: 43.1 % ± 15.7, MC1: 54.1 % ± 16.7, p=0.018) (Figure 3c).

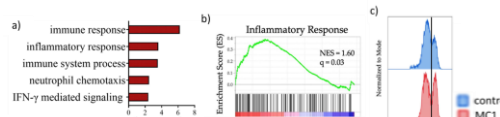


Figure 3. Transcriptomic analysis of MC1 and intra-patient control bone marrow neutrophils. a) Gene ontology overrepresentation analysis: Top 5 upregulated biological processes in MC1 compared to intra-patient control bone marrow neutrophils. b) Gene set enrichment analysis: Enriched "inflammatory" gene set in MC1 bone marrow neutrophils. c) Representative flow cytometry image of CD66b expression in MC1 and control bone marrow neutrophils.

## Discussion

- MC1 bone marrow neutrophils have an activated phenotype
- Activated peripheral blood neutrophils degrade proteoglycans from CEPs
- sGAG released from CEPs exposed to activated neutrophil supernatant for only 18 hours is similar to the amount of sGAG lost *in vivo* over 20 years of natural ageing
- Whether activated MC1 bone marrow neutrophils can degrade CEP tissue remains to be elucidated

### 2. Activated blood neutrophils degrade human cartilage endplates (CEPs)

#### Exposure of CEP tissues to conditioned medium from activated neutrophils:

- Significant increased sGAG release from CEP tissues in a dose-dependent manner (Figure 4)
  - 25 mio/ml: 380.1 % ± 177, p = 0.012;
  - 12.5 mio/ml: 123.7 % ± 22.3, p = 0.048
- Relative sGAG release: 3.1-fold higher in the CEPs exposed to supernatant from 25 mio/ml neutrophils compared to 12.5 mio/ml neutrophils (25 vs. 12.5 mio/ml: p = 0.022)
- No significant effect of neutrophil supernatant on hydroxyproline release

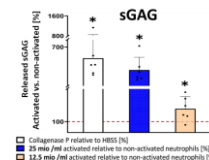


Figure 4. Neutrophil-mediated CEP damage model. Relative sGAG released from CEP tissues exposed to positive control (white bars) or neutrophil supernatant from 25 mio/ml (blue) and 12.5 mio/ml blood neutrophils (brown). sGAG release from half-biopsy specific control was set to 100 %. \*P < 0.01.

## Implications

Activated MC1 bone marrow neutrophils may promote and exacerbate CEP damage, thereby facilitating enhanced inflammatory disc/marrow crosstalk in MC1.

CEP damage may also increase nerve fibre density.



These findings reveal a potential novel MC1 pathomechanism and could have implications for treatment strategies to mitigate CEP damage in MC1.